# Toxicant default guideline values for aquatic ecosystem protection

Metsulfuron-methyl in freshwater

Technical brief

July 2021

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## Summary

The default guideline values (DGVs) and associated information in this technical brief should be used in accordance with the detailed guidance provided in the Australian and New Zealand Guidelines for Fresh and Marine Water Quality website (www.waterquality.gov.au/anz-guidelines).

Metsulfuron-methyl (2-(4-methoxy-6-methyl-1,3,5-triazin-2-ylcarbamoylsulfamoyl)benzoate, CAS no. 74223-64-6) is a selective, systemic triazinylsulfonylurea herbicide that sits within the sulfonylurea group of the urea family of herbicides. Other triazinylsulfonylurea herbicides include iodosulfuron, ethametsulfuron, thifensulfuron and their methylated forms. In Australia and New Zealand, metsulfuron-methyl is typically used in agriculture to control broadleaf and other weeds in pastures, cereals, forestry and/or for other non-agricultural purposes (ACVM 2020, APVMA 2020). Metsulfuron-methyl exerts toxicity by binding to and inhibiting the acetolactate synthase (ALS) enzyme, which prevents the biosynthesis of amino acids and stops cell division. Non-agricultural uses of metsulfuron-methyl include weed control in forestry.

The previous Australian and New Zealand DGV for metsulfuron-methyl in freshwater environments was a low reliability value (using the ANZECC/ARMCANZ (2000) reliability scheme) as it was based on acute toxicity data for two freshwater fish species, one invertebrate and one microalga only (ANZECC/ARMCANZ 2000). More data on metsulfuron-methyl toxicity to freshwater species are now available, including data on phototrophic species (species that photosynthesise e.g. plants and algae), resulting in moderate reliability DGVs (Warne et al. 2018).

Due to the small sample size of available heterotrophic ecotoxicity data for metsulfuron-methyl, it was not possible to ascertain distinctions in sensitivity between different groups of species, for example between phototrophic and heterotrophic species. As a result, both phototrophic and heterotrophic species were used to derive the metsulfuron-methyl DGVs, as recommended in Warne et al. (2018). However, based on the mode of action of metsulfuron-methyl, it is expected that phototrophic species would be more sensitive than non-phototrophic species, as metsulfuron-methyl binds to and inhibits the ALS enzyme, which is a protein generally found in plants. Consequently, it is possible that the derived DGVs for metsulfuron-methyl may not provide adequate protection to all phototrophic species. In addition to this, as phototrophs are at the bottom of most aquatic food webs, the derived DGVs may not provide sufficient protection to non-phototrophic species (as a result of potential indirect effects).

The lowest reported chronic toxicity value to freshwater species is 0.054 µg/L (freshwater macrophyte, 8-day NOEC). The lowest reported acute toxicity value to freshwater species is 64.8 µg/L (freshwater microalga, 24-hour EC5).

The DGV derivation for metsulfuron methyl in freshwater incorporated chronic EC10, NOEC and NOEL data for eight freshwater, phototrophic and heterotrophic, species from five phyla and six classes. The (visual) fit of the model to the toxicity data was poor, resulting in moderate reliability DGVs. The DGVs derived here are expressed in terms of the active ingredient (metsulfuron-methyl) rather than commercial formulations. The DGVs for 99, 95, 90 and 80% species protection are 0.0037 µg/L, 0.018 µg/L, 0.048 µg/L and 0.18 µg/L, respectively. The 95%species protection level for metsulfuron-methyl in freshwater (0.018 µg/L) is recommended for adoption in the assessment of slightly-to-moderately disturbed ecosystems.

## Introduction

Metsulfuron-methyl is a herbicide (C14H15N5O6S; see Figure 1) that at room temperature is a white to pale-yellow solid with a characteristic ester-like odour. It is the active ingredient of a variety of commercial herbicide formulations. Metsulfuron-methyl is often recommended to be mixed with other herbicides (e.g. terbutryn and glyphosate formulations) in tank mixes to increase its efficacy. Physico-chemical properties of metsulfuron-methyl that may affect its environmental fate and toxicity are presented in Table 1.

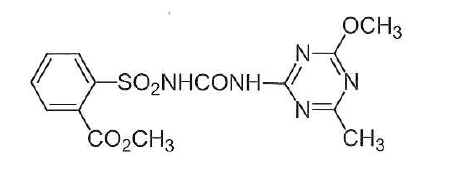


Figure 1 Structure of metsulfuron-methyl

Table 1 Summary of selected physico-chemical properties of metsulfuron-methyl

| Physico-chemical property | Value |
| --- | --- |
| Molecular weight | 381.4 amu |
| Aqueous solubility | 548 mg/L (pH 5), 2 790 mg/L (pH 7), 213 000 mg/L (pH 9) @ temperature of 25 oC **1**2 790 mg/L @ temperature of 20 oC **2** |
| Logarithm of the octanol-water partition coefficient (log Kow) | -1.87 at pH 7 and temperature of 20 oC **a**, **b** |
| Logarithm of the organic carbon water partition coefficient (log Koc) | 1.54 **c** |
| Logarithm of the bioconcentration factor (log BCF) | 1 **c** |
| Half-life (t1/2) in water | 22 days at pH 5–9 and temperature of 25 ºC **b** |
| Half-life (t1/2) in soil | Typical: 10–30 days; up to 180 days **b**, **d** |

**a** BCPC (2012).

**b** Pesticide Properties Database (University of Hertfordshire 2013).

**c** Barcelö and Hennion (2003).

**d** Cornell University (1993).

Metsulfuron-methyl belongs to the triazinylsulfonylurea group that sits within the sulfonylurea group of the urea family of herbicides, which also includes iodosulfuron, ethametsulfuron, thifensulfuron and their methylated forms. It is a systemic herbicide and can be applied before and/or after weeds emerge (i.e. it is a pre-emergent and post-emergent herbicide). In Australia, metsulfuron-methyl has been approved for brush and broadleaf weed control on a range of crops including cereals (e.g. wheat, barley, canola, rye, triticale), linseed, chickpeas, mung beans and pastures, and also in other activities such as forestry, commercial and industrial areas, and for *Mimosa pigra* control on floodplains (APVMA 2020). In New Zealand, it is approved for weed control on pastures, and in forestry (radiata pine) and non-cropland areas (ACVM 2020).

Metsulfuron-methyl is a selective residual herbicide, and it retains its biological effectiveness in soil, with a half-life ranging from days to months (Cornell University 1993, University of Hertfordshire 2013) and recommended exclusion times for some crops of up to 22 months (Cornell University 1993). The higher the soil moisture content and temperature and the lower the acidity, the more rapidly metsulfuron-methyl is degraded in soil (Smith 1986).

Metsulfuron-methyl has a low affinity for binding to soil particles (Table 1); therefore, it has a high capacity to leach to groundwater and end up in surface waters. The aqueous hydrolysis of metsulfuron-methyl is relatively fast, with a half-life of 22 days at pH 5–9 and a temperature of 25 ˚C (University of Hertfordshire 2013) (Table 1).

## Aquatic toxicology

### Mechanism of toxicity

Metsulfuron-methyl is mainly absorbed through the roots and foliage of plants and is transported to the leaves and shoots where it exerts its toxicity. Metsulfuron-methyl binds to, and inhibits, the acetolactate synthase (ALS) enzyme, which is responsible for catalysing the formation of amino acids. As a result, the biosynthesis of amino acid branches within sensitive plants is inhibited (FAO UN 2015). Exposed plants typically die within two to four weeks due to cessation of cell division and growth processes.

### Toxicity

#### Freshwater chronic toxicity data

There were freshwater chronic toxicity data for one fish, 12 macrophytes and six microalgae.

The toxicity values for the single fish species were:

* 90-day NOEL and LOEC values of 4 500 µg/Land 8 000 µg/L.

The toxicity values for macrophytes consisted of:

* a 6-day EC50 (dry weight) value of 11.95 µg/L
* 7-day EC10 (frond count and frond area) values ranging from 0.11 µg/L to 1.45 µg/L
* 7-day EC50 (frond number and frond area) values ranging from 0.06 µg/L to 12.7 µg/L
* 8-day NOEC (dry weight, shoot length and root growth) values ranging from 0.054 µg/L to 20 µg/L
* 8-day LOEC (shoot length) values ranging from 0.1 µg/L to 1 µg/L
* a 9-day EC50 (dry weight) value of 6.18 µg/L
* an 11-day EC50 (frond number) value of 0.68 µg/L
* a 12-day EC50 (dry weight) value of 2.61 µg/L
* 14-day NOEC/NOEL (root occurrence, dry weight, frond number and frond area) values ranging from 0.16 µg/L to 20 µg/L
* 14-day LOEC (chlorophyll-a content and root occurrence) values ranging from 1 µg/L to 2.7 µg/L
* 14-day EC50 (leaf area, fresh weight: dry weight, frond number, frond area, chlorophyll-a content and total shoot length) values ranging from 0.1 µg/L to 26.7 µg/L
* a 15-day EC50 (dry weight) value of 2.43 µg/L
* 42-day NOEC, LOEC and EC50 (frond count) values of 0.1 µg/L, 0.5 µg/L and 0.99 µg/L, respectively.

The toxicity values for microalgae were:

* a 48-hour EC10 (chlorophyll-a content) value of 292 µg/L
* two 48-hour EC50 (chlorophyll-a content) values of 677 µg/L and 1 934 µg/L
* a 72-hour IC50 (cell density) value of 612 µg/L
* two 96-hour NOEL (biomass yield, growth rate and area under the growth curve) values of 14.5 µg/L and 92 800 µg/L
* 96-hour EC50 (cell density, biomass, growth rate and area under the growth curve) values ranging from 130 µg/L to 72 782 µg/L
* two 5-day NOEL (biomass, growth rate and area under the curve) values of 10 µg/L and 95.4 µg/L
* a 5-day EC50 (biomass, growth rate and area under the curve) value of 286 µg/L
* 6-day EC20 and EC50 (chlorophyll-a content) values of 68.6 µg/L and 1 564 µg/L, respectively.

#### Freshwater acute toxicity data

There were freshwater acute toxicity data for one microalga and one macrophyte.

The toxicity values for the single microalga species were:

* 24-hour NOEC, EC5 and EC50 (cell density) values of 165.1 µg/L, 64.8 µg/L and 1 163 µg/L, respectively.

The toxicity values for the single macrophyte species were:

* 96-hour LOEC and EC50 (frond count) values of 0.2 µg/L and 0.4 µg/L, respectively.

As stated in Warne et al. (2018), acute EC10/NOEC and LOEC values should not be converted to chronic EC10/NOEC values, and these have not been used to derive DGVs.

## Factors affecting toxicity

No factors have been reported as modifying the toxicity of metsulfuron-methyl. As with many organic chemicals, it might be expected that dissolved and particulate organic matter and suspended solids would affect its bioavailability and toxicity. Kah and Brown (2007) found that sorption to soil was negatively correlated to soil pH and positively correlated to soil organic carbon, such that relatively strong sorption was measured in soils with low pH and high organic carbon content. However, overall, sorption of metsulfuron-methyl to soil was still relatively weak compared to many other pesticides (Kah and Brown 2007), which is supported by its low organic carbon water partition coefficient (log Koc) (Table 1).

## Default guideline value derivation

The DGVs were derived in accordance with the method described in Warne et al. (2018) and using Burrlioz 2.0 software. Although some decisions on data selection/manipulation may reflect the Warne et al. (2015) method rather than the Warne et al. (2018) method, these were found to have no material effect on the final DGVs.

### Toxicity data used in derivation

The previous Australian and New Zealand Guidelines for Fresh and Marine Water Quality (ANZECC/ARMCANZ 2000) included an environmental concern level (ECL) for metsulfuron-methyl in freshwater environments. This ECL had a low reliability based on the ANZECC/ARMCANZ (2000) reliability scheme as it was based on unscreened acute toxicity data for only two freshwater fish, one invertebrate and one microalga (ANZECC/ARMCANZ 2000).

To obtain toxicity data for metsulfuron-methyl for freshwater organisms, an extensive search of the scientific literature was conducted. In addition, the ECOTOXicology Database System (USEPA 2015a), Office of Pesticide Programs Database (USEPA 2015b), the Australasian Ecotoxicology Database (Warne et al. 1998) and the ANZECC/ARMCANZ (2000) toxicant database (Sunderam et al. 2000) were searched. There are now considerably more metsulfuron-methyl toxicity data available to enable the calculation of DGVs for freshwater (Appendix A: Toxicity data that passed the screening and quality assessment and were used to derive the default guideline values ). However, as there are still insufficient chronic no or low effect toxicity data to meet the preferred dataset size (i.e. toxicity data for at least 15 species that belong to at least four taxonomic groups), it is recommended that additional chronic toxicity tests of metsulfuron-methyl with phototrophic (e.g. plants and algae) freshwater species be conducted.

In total, there were freshwater toxicity data for 20 species (five phyla and seven classes) that passed the screening and quality assessment processes. The represented phyla were Bacillariophyta, Chlorophyta, Chordata, Cyanobacteria and Tracheophyta. The seven classes were Actinopterygii (which accounts for approximately 99% of fish), Bacillariophyceae (diatoms; a major grouping of algae), Chlorophyceae (a major grouping of freshwater green algae), Cyanophyceae (a class of cyanobacteria), Liliopsida (monocots), Magnoliopsida (dicots) and Trebouxiophyceae (another grouping of green algae). Chronic toxicity data were available for 19 of the 20 species, comprising 18 phototrophs and one heterotroph, while only acute toxicity data were available for one species—a phototroph.

Based on the current understanding of the mode of action of metsulfuron-methyl, an ALS-inhibiting herbicide, it is expected that phototrophic species would be more sensitive than non-phototrophic species. However, due to the small sample size of heterotrophic species, it was not possible to discern differences in sensitivity between different groups of species, for example between phototrophic and heterotrophic species. Therefore, both phototrophic and heterotrophic species were used to derive the metsulfuron-methyl DGVs, as recommended in Warne et al. (2018).

From the dataset of toxicity values for 20 species, there were freshwater chronic no or low effect (i.e. EC10, NOEC, NOEL, LOEC) data for eight species (seven phototrophic and one heterotrophic) that belonged to five phyla. This met the minimum data requirements necessary to use a species sensitivity distribution (SSD) to derive DGVs. However, the fit of the SSD to the data was poor and, combined with the number and type of toxicity data, this resulted in the DGVs having moderate reliability. In an attempt to improve the reliability of the DGVs, chronic EC50 type data (for the remaining 12 species; converted to no or low effect estimates by dividing by 5, as per Warne et al. 2018) were added to the dataset. However, the resulting SSD fitted this dataset worse than the SSD for the chronic no or low effect dataset, particularly for the more sensitive species. Therefore, the DGVs for metsulfuron-methyl were derived using only the chronic no or low effect data.

To identify species that were regionally relevant to Australia and New Zealand ecosystems, a search of AlgaeBase (Guiry & Guiry 2017), Atlas of Living Australia (ALA 2017), Catalogue of Life (Roskov et al. 2017), Integrated Taxonomic Information System (ITIS 2017) and the World Register of Marine Species (WoRMS 2017) was conducted. The dataset used in the DGV derivation process for metsulfuron-methyl in freshwater includes toxicity data for four freshwater species that either originated from or are distributed within Australia and/or New Zealand.

A summary of the toxicity data used to calculate the DGVs for metsulfuron-methyl in freshwater is provided in Table 2. A summary of the acceptable quality raw toxicity data for all freshwater species that passed the screening and quality assurance processes is provided in Section 2.2 and Section 2.2.2. Further details on the data that passed the quality assessment and screening process and were used to derive the DGVs are presented in Appendix A: Toxicity data that passed the screening and quality assessment and were used to derive the default guideline values, Table A 1. Details of the data quality assessment and the data that passed the quality assessment are provided as supporting information.

Table 2 Summary of the single toxicity values, all species used to derive default guideline values for metsulfuron-methyl in freshwater

| Taxonomic group (Phylum) | Species | Life stage | Duration (d) | Type (acute/ chronic) | Toxicity endpoint | Final toxicity value (µg/L) |
| --- | --- | --- | --- | --- | --- | --- |
| Blue–green alga (Cyanobacteria) | Anabaena flosaquae | Not stated | 5 | Chronic NOEL | Biomass yield, growth rate, AUC **a** | 95.4 |
| Macrophyte (Tracheophyta) | Elodea canadensis**b** | Apical shoot (19.5 cm) | 8 | Chronic NOEC | Shoot length | 0.054 |
| Macrophyte (Tracheophyta) | Lemna gibba | Not stated | 7 | Chronic EC10 | Frond count | 0.193 c |
| Macrophyte (Tracheophyta) | Lemna minor**b** | Exponential growth phase | 42 | Chronic NOEC | Frond count | 0.1 |
| Macrophyte (Tracheophyta) | Myriophyllum spicatum | Apical shoot | 14 | Chronic LOEC | Root occurrence | 0.4 **d** |
| Diatom (Bacillariophyta) | Navicula pelliculosa **b** | Not stated | 4 | Chronic NOEL | Biomass yield, growth rate, AUC **a** | 92 800 |
| Fish (Chordata) | Oncorhynchus mykiss**b** | Early life | 90 | Chronic NOEL | Mortality | 4 500 |
| Green alga (Chlorophyta) | Pseudokirchneriella subcapitata**e** | Not stated | 5 | Chronic NOEL | Biomass yield, growth rate, AUC **a** | 10 |

**a** AUC = area under the growth curve.

**b** Species that originated from, or whose geographic distributions include, Australia and/or New Zealand.

**c** Represents a geometric mean of four toxicity values.

**d** Converted to an estimated chronic NOEC/EC10 by dividing by 2.5 (Warne et al. 2018).

**e** This species has also been called Raphidocelis subcapitata and Selenastrum capricornutum.

### Species sensitivity distribution

The cumulative frequency (species sensitivity) distribution (SSD) of the eight freshwater phototrophic and heterotrophic species that was used to derive the DGVs is presented in Figure 2. The SSD was plotted using the Burrlioz 2.0 software, and the model was judged to provide a poor fit to the data.

Figure shows the species sensitivity distribution of several species at different concentrations of metsulfuron-methyl. Chronic NOEL, chronic NOEC and chronic EC10 values are shown on the figure.

Figure 2 Species sensitivity distribution, metsulfuron-methyl in freshwater

### Default guideline values

It is important that the DGVs (Table 3) and associated information in this technical brief are used in accordance with the detailed guidance provided in the Australian and New Zealand Guidelines for Fresh and Marine Water Quality website. As with other pesticides that have DGVs, the DGVs for metsulfuron-methyl are expressed in terms of the concentration of the active ingredient.

The derived Australian and New Zealand DGVs for metsulfuron-methyl in freshwater are provided in Table 3. The 99% and 95% species protection DGVs may be below routine limits of reporting for most commercial laboratories, although some laboratories may be able to achieve lower limits of reporting. ANZG (2018; see [Accounting for local conditions](https://www.waterquality.gov.au/anz-guidelines/guideline-values/default/water-quality-toxicants/local-conditions#default-guideline-values-and-analytical-detection-limits)) provides guidance on what to do if DGVs are below analytical detection limits. By including phototrophic and heterotrophic species in the dataset used to derive DGVs for metsulfuron-methyl, which is expected to be more toxic to phototrophs, it is possible that the DGVs may not provide adequate protection to all phototrophic species. Additionally, as phototrophs are at the bottom of most aquatic food webs, the DGVs also may not provide sufficient protection to non-phototrophic species, as a result of potential indirect effects.

Measured log BCF values for metsulfuron-methyl are low (Table 1) and are below the threshold at which secondary poisoning must be considered (i.e. threshold log BCF = 4, Warne et al. 2018). Therefore, the DGVs for metsulfuron-methyl do not need to account for secondary poisoning.

Table 3 Toxicant default guideline values, metsulfuron-methyl in freshwater, moderate reliability

| Level of species protection (%) | DGV for metsulfuron-methyl in freshwater (µg/L) ****a**** |
| --- | --- |
| 99 | 0.0037 |
| 95 | 0.018 |
| 90 | 0.048 |
| 80 | 0.18 |

**a** DGVs were derived using the Burrlioz 2.0 software and rounded to two significant figures.

### Reliability classification

The metsulfuron-methyl in freshwater DGVs have a moderate reliability classification (Warne et al. 2018) based on the outcomes for the following three criteria:

* Sample size—8 (good)
* Type of toxicity data—chronic EC10, NOEC, LOEC
* SSD model fit—poor (Inverse Weibull model).

## Glossary

| Term | Definition |
| --- | --- |
| acute toxicity | A lethal or adverse sub-lethal effect that occurs as the result of a short exposure period to a chemical relative to the organism’s life span. |
| ANZECC | Australian and New Zealand Environment and Conservation Council |
| ARMCANZ | Agricultural and Resource Management Council of Australia and New Zealand |
| CAS no. (Chemical Abstracts Service number) | Each chemical has a unique identifying number that is allocated to it by the American Chemical Society. |
| chronic toxicity | A lethal or sublethal adverse effect that occurs after exposure to a chemical for a period of time that is a substantial portion of the organism’s life span or an adverse effect on a sensitive early life stage. |
| default guideline value (DGV) | A guideline value recommended for generic application in the absence of a more specific guideline value (e.g. site-specific guideline value), in the Australian and New Zealand Guidelines for Fresh and Marine Water Quality. Formerly known as ‘trigger values’. |
| EC50 (median effective concentration) | The concentration of a substance in water or sediment that is estimated to produce a 50% change in the response being measured or a certain effect in 50% of the test organisms relative to the control response, under specified conditions. |
| ECx | The concentration of a substance in water or sediment that is estimated to produce an x% change in the response being measured or a certain effect in x% of the test organisms, under specified conditions. |
| endpoint | The specific response of an organism that is measured in a toxicity test (e.g. mortality, growth, a particular biomarker). |
| environmental fate | The behaviour (e.g. movement, transformation, degradation, partitioning) of a chemical in the environment. |
| guideline value (GV) | A measurable quantity (e.g. concentration) or condition of an indicator for a specific community value below which (or above which, in the case of stressors such as pH, dissolved oxygen and many biodiversity responses) there is considered to be a low risk of unacceptable effects occurring to that community value. Guideline values for more than one indicator should be used simultaneously in a multiple lines of evidence approach. (Also refer to ‘default guideline value’ and ‘site-specific guideline value’.) |
| ICx | The concentration of a substance in water or sediment that is estimated to produce an x% inhibition in the response being measured relative to the control response, under specified conditions. |
| LC50 (median lethal concentration) | The concentration of a substance in water or sediment that is estimated to be lethal to 50% of a group of test organisms, relative to the control response, under specified conditions. |
| lowest observed effect concentration (LOEC) | The lowest concentration of a material used in a toxicity test that has a statistically significant adverse effect on the exposed population of test organisms as compared to the controls. |
| Macrophyte | A member of the macroscopic plant life of an area, especially of a body of water; large aquatic plant. |
| no observed effect concentration (NOEC)  or  no observed effect level (NOEL) | The highest concentration of a material used in a toxicity test that has no statistically significant adverse effect on the exposed population of test organisms as compared with the controls. |
| phototrophs | Organisms that photosynthesize as their main means of obtaining energy e.g. plants and algae. |
| site-specific guideline value | A guideline value that is relevant to the specific location or conditions that are the focus of a given assessment or issue. |
| Species (biological) | A group of organisms that resemble each other to a greater degree than members of other groups and that form a reproductively isolated group that will not produce viable offspring if bred with members of another group. |
| species sensitivity distribution (SSD) | A method that plots the cumulative frequency of species’ sensitivities to a toxicant and fits a statistical distribution to the data. From the distribution, the concentration that should theoretically protect a selected percentage of species can be determined. |
| toxicity | The inherent potential or capacity of a material to cause adverse effects in a living organism. |
| toxicity test | The means by which the toxicity of a chemical or other test material is determined. A toxicity test is used to measure the degree of response produced by exposure to a specific level of stimulus (or concentration of chemical) for a specified test period. |

## Appendix A: Toxicity data that passed the screening and quality assessment and were used to derive the default guideline values

Table A 1 Summary, chronic toxicity data that passed the screening and quality assurance processes, metsulfuron-methyl in freshwater

| Taxonomic group (Phylum) | Species | Life stage | Exposure duration (d) | Toxicity measure (test endpoint) | Test medium | Temperature (°C) | pH | Concentration (µg/L) | Reference |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Diatom  (Bacillariophyta) | Navicula pelliculosa | Not stated | 4 | NOEL  (Biomass, growth rate, AUC **b**) | ASTM Type I water | 24 ± 2 | 7.5 ± 0.1 | 92 800 | USEPA (2015b) |
| – | | | | | | | | **92 800** | **VALUE USED IN SSD** |
| Green alga  (Chlorophyta) | Selenastrum capricornutum**a** | Not stated | 5 | NOEL  (Biomass, growth rate, AUC **b**) | ASTM Type I water | 24 ± 2 | 7.5 ± 0.1 | 10 | USEPA (2015b) |
| – | | | | | | | | **10** | **VALUE USED IN SSD** |
| Fish  (Chordata) | Oncorhynchus mykiss | Early life | 90 | NOEL  (Mortality) | Dilution water | 12 ± 2 | Not stated | 4 500 | USEPA (2015b) |
| – | | | | | | | | **4 500** | **VALUE USED IN SSD** |
| Blue–green alga  (Cyanobacteria) | Anabaena flosaquae | Not stated | 5 | NOEL  (Biomass, growth rate, AUC **b**) | ASTM Type I water | 24 ± 2 | 7.5 ± 0.1 | 95.4 | USEPA (2015b) |
| – | | | | | | | | **95.4** | **VALUE USED IN SSD** |
| Macrophyte  (Tracheophyta) | Elodea canadensis | Apical shoot (19.5 cm) | 8 | NOEC  (Shoot length) | Filtered freshwater | 22 ± 2 | Not stated | 0.054 | Wendt-Rasch et al. (2003) |
| – | | | | | | | | **0.054** | **VALUE USED IN SSD** |
| Macrophyte  (Tracheophyta) | Lemna gibba | Not stated | 7 | EC10  (Frond count) | 20 × AAP medium | 24 ± 2 | 7.5 | 0.27 | Rosenkrantz et al. (2013) |
| Not stated | 7 | EC10  (Frond count) | 20 × AAP medium | 24 ± 2 | 7.5 | 0.16 | Rosenkrantz et al. (2013) |
| Not stated | 7 | EC10  (Frond count) | 20 × AAP medium | 24 ± 2 | 6.0 | 0.12 | Rosenkrantz et al. (2013) |
| Not stated | 7 | EC10  (Frond count) | 20 × AAP medium | 24 ± 2 | 9.0 | 0.27 | Rosenkrantz et al. (2013) |
| – | | | | | | | | **0.193** | **VALUE USED IN SSD (GEOMETRIC MEAN)** |
| Macrophyte  (Tracheophyta) | Lemna minor | Exponential growth phase | 42 | NOEC  (Frond count) | Swedish standard (SIS) | 20 ± 1 | 6.5 ± 0.2 | 0.1 | Boxall et al. (2013) |
| – | | | | | | | | **0.1** | **VALUE USED IN SSD** |
| Macrophyte  (Tracheophyta) | Myriophyllum spicatum | Apical shoot | 14 | LOEC  (Root occurrence) | Filtered freshwater | 22 ± 2 | Not stated | 1.0 | Wendt-Rasch et al. (2003) |
| – | | | | | | | | **0.4 c** | **VALUE USED IN SSD** |

**a** This species has been called Raphidocelis subcapitata, Pseudokirchneriella subcapitata and Selenastrum capricornutum.

**b** AUC = area under the growth curve.

**c** LOEC converted to an estimated chronic NOEC/EC10 by dividing by 2.5 (Warne et al. 2018).

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